



## RESEARCH ARTICLE

## Use of an Antigen Excreted (SODe) in the Search for Antibodies Anti-*Trypanosoma Cruzi* in Sera from Pediatric Population of the State of Queretaro, Mexico

María Elena Villagrán-Herrera<sup>1\*</sup>, Manuel Sánchez-Moreno<sup>2</sup>, José Alejandro Martínez-Ibarra<sup>3</sup>, Ricardo Francisco Mercado-Curiel<sup>1</sup>, Adriana Jheny Rodríguez-Méndez<sup>1</sup>, Javier Ávila-Morales<sup>1</sup>, María del Carmen Aburto-Fernández<sup>1</sup>, Nicolás Camacho-Calderón<sup>1</sup> and José Antonio de Diego-Cabrera<sup>4</sup>



<sup>1</sup>Departamento de Investigación Biomédica, Facultad de Medicina, Universidad Autónoma de Querétaro, México

<sup>2</sup>Departamento de Parasitología Molecular, Facultad de Ciencias, Universidad de Granada, España

<sup>3</sup>Área de Entomología Médica, Centro Universitario del Sur, Universidad de Guadalajara 49000, México

<sup>4</sup>Unidad de Parasitología y Medicina Tropical, Departamento de Medicina Preventiva, Salud Pública y Microbiología, Facultad de Medicina, Universidad Autónoma de Madrid, España

\*Corresponding author: María Elena Villagrán Herrera, Departamento de Investigación Biomédica, Facultad de Medicina, Universidad Autónoma de Querétaro, Clavel No. 200, Fraccionamiento la Capilla, C.P. 76170, Santiago de Querétaro, México, Tel: +52-442-1921200, ext. 62550, E mail: [mevh@uaq.mx](mailto:mevh@uaq.mx)

### Abstract

The acute phase of Chagas disease can occur at any age, however the pediatric population is usually the most affected (from one to 15 years), so it is very important to detect and treat it as soon as possible to try to eliminate the circulating parasite.

**Objective:** The main objective of this research is to present the first seroepidemiological study of the infection by *Trypanosoma cruzi*, carried out to students of school age of both urban and rural primary schools in the State of Querétaro and at the same time, the validation of an enzyme excreted by the hemoflagellate parasite, Superoxide Dismutase (SODe), as an adequate molecular marker.

**Methods:** In this study, a total of 540 sera collected between February 2015 and May 2016 were evaluated through ELISA and Western blot tests using the marker excreted as an antigenic part, in an ELISA-SODe and a WB-SODe.

**Results:** Of the total of sera studied 106 (19.6%) gave reactivity. Of these, 99 sera were reactive against ELISA-SODe (prevalence of 18.3%), whereas 100 sera were reactive for WB-SODe (prevalence of 18.52%). The results of the two applied tests were evaluated and compared, using the Western blot technique, as gold standard, to find that the sensitivity and specificity of ELISA-SODe reach respectively 93% and 99%.

**Conclusions:** Our study provides seroepidemiological data on the presence of chagasic infection in the pediatric population of rural and urban areas in Santiago de Querétaro and, at the same time, we confirm that the SODe excreted by *Trypanosoma cruzi* is highly immunogenic, which indicates it as an excellent tool for the early diagnosis of Chagas disease.

### Keywords

*Trypanosoma cruzi*, Superoxide dismutase, Immunogenic, Pediatric population, Rural, Urban

### Introduction

American trypanosomiasis is recognized by WHO as one of the 17 neglected tropical diseases in the world. Caused by the hemoflagellate parasite *Trypanosoma cruzi*, which in its natural form is transmitted by vectors of the order *Hemiptera* (Bugs), family *Reduviidae*, subfamily, *triatominae* and *triatoma* genus, and by non-vector mechanisms, such as blood transfusions, organ transplants, transmission orally and vertically from mother to child [1,2].

The Pan American Health Organization [3] estimates that there are currently 7.7 million people infected with *T. cruzi* in 21 endemic countries, with 41,200 new cases per year, and 14,400 children born annually with congenital Chagas disease. The most affected countries are: Bolivia (6.8%), Argentina (4.1%), El Salvador (3.4%), Honduras (3.1%), Paraguay (2.5%), Guatemala (2%), Ecuador (1.7%), French Guyana, Guyana, Suriname (1.2%), Venezuela (1.2%), Nicaragua (1.1%) and Brazil (1%), Panama, Peru, Mexico, Belize, Venezuela, Costa Rica, Chile and Paraguay [4].

Mexico, according to the official figures of the World Health Organization (WHO), estimates that 1.1 million people are infected with the trypanosome, and according to Hotez, et al. [5] in their research has recorded an estimate of 1-6 million infected.

The disease is considered a serious public health problem mainly in rural areas of Mexico, Central and South America. Migration has been an important factor, since there are currently cases reported from Latin America, which have been detected in the US, Europe, Japan, Australia. This is in addition to the enzootic transmission of *Trypanosoma cruzi*, which involves several species of mammals and vectors, some of them mention raccoons, opossums and especially domestic dogs [5,6].

Therefore, it should be considered that the acquisition of the trypanosomatid infection is not limited to rural areas, since in México, Peru, Bolivia and Venezuela cases have been reported in urban and peri-urban areas [7].

The State of Querétaro is not officially considered as an endemic area of the infection; The obligatory tests of ELISA are carried out only in blood samples from patients coming from risk areas and in reactive cases, confirmatory tests are not carried out. The above, added to the central location of the state of Querétaro, leads to a constant leakage of individuals from other states, previously transfused, or infected, ignoring whether their blood is contaminated or not, they are offered for blood donors. In 2005, the State Center for Blood Transfusion (CETS) reported 0.3% of HIV-positive *Trypanosoma cruzi* in its blood donors. By not having reliable statistical data to help quantify the real situation of endemic areas, Villagrán and collaborators in that same year, began a seroepidemiological cross-sectional study in 23 communities of 11 rural municipalities in the State of Querétaro. Collecting 1029 blood samples from the general population to which 4 conventional tests were applied, obtaining a seroprevalence of 6.2%, to two reactive tests, which compared to the national average of 1.6% obtained by Guzman-Bracho, et al. [8] in 2001, it exceeded by 5 times the percentage reported and by far the data contributed by the state CETS of 0.3%.

Today, the main form of infection in the rural area is not vector transmission, since the congenital transmission (transplacental), has far exceeded the bite of the bug. In Mexico, the first case of neonatal Chagas was reported in 1998 and the following case was reported in 2012 by Jiménez Cardoso, et al. [9] who detected an average of 4.08% of specific antibodies against *T. cruzi* in infants of seropositive mothers in states such as Oaxaca and Jalisco.

The diagnosis of Chagas disease depends on the stage in which the disease is present in the patient and can be performed by direct techniques (visualization of the parasite in the peripheral blood), xenodiagnosis or indirect (immunological tests), and the reaction in polymerase chain. (PCR) as a last resort, 100% sensitive and specific. Due to the high costs of PCR, it is not possible to keep it viable in most clinical laboratories, in addition, the lack of protocols to standardize, the best products to amplify, the sensitivity can vary from 45% to 96.5%, therefore, false positives can be obtained [10-12]. On the other hand, the WHO [13], states that two serological tests must be performed other than the serum of a probable Chagasic case, if both are reactive, the individual is considered positive, however if one of two is negative, you must perform a third immunological test. The above reflects that the diagnosis of this disease is expensive and is beyond the reach of the population that suffers the greatest risk of acquiring it, so it is important and necessary to seek and find new reliable diagnostic tools. Many studies tend to define a specific antigen of *T. cruzi* that would increase the specificity and sensitivity of the serodiagnosis. A possible candidate could be the Superoxide Dismutase of iron excreted by *T. cruzi* (Fe-SODe or SODe), which, in previous studies, has shown to have highly specific immunogenic properties, which make it a useful, unique tool to diagnose the disease in a population from or not from an endemic area [14-17].

## Material and Methods

### Parasites and crops

The epimastigote forms of *Trypanosoma cruzi* (MHOM/ME/2006/H-4) were cultured in axenic medium. Liquid medium for trypanosomes (MTL, Gibco) supplemented with 10% heat inactivated bovine fetal serum at 28 °C in Falcon flasks according to Longoni, et al. [18] was used.

### Selection of participants and study design

Random visits were made in several primary schools in three rural areas and two in the urban area of the State of Querétaro (Mexico).

Being a cross-sectional and longitudinal study, we are guided by the seroprevalence obtained in previous investigations with the general and adult population, in the chosen areas. Public elementary schools were chosen from localities already studied and considered

endemic, in addition to meeting the pediatric age limit of 18 years (According to the Ministry of Health). The minimum number of school population in each school is 200 children, and maximum of 500, so that the N real (number of real samples) of work, will be the number of surveys with the informed consent delivered and signed by the parents of family who authorize the study to be carried out on their children. The statistical program Epi-info version 2, Statcalc module, was of great help, considering an expected serological prevalence of 6.2%, for the study conducted in 2009 in rural communities and an unexpected frequency of 9%.

Initial interviews were scheduled with school principals and parents of children, where information about the importance of the study was presented. Epidemiological surveys and informed consent signed by parents authorizing research in their children and filling out the surveys with all the relevant data of their children before their examination.

The number of blood samples per rural location was as follows: Community of El Gallo, 77, (1 to 77), San Vicente Ferrer, 106, (78 to 183) and San Ignacio, 111, (335 to 445), each one of them with 1,839, 1,525 and 871 inhabitants respectively. The selected communities belong to the municipalities of El Marqués, Colón and Huimilpan, respectively. Regarding the urban area of the municipality of Querétaro, the total blood samples obtained were 244, (184 to 540). This municipality has a population of 626,517 inhabitants.

A total of 540 sera (collected from February 2015 to May 2016) were evaluated, which were not grouped in any classification category (they were only numbered from 1 to 540).

The protocol designed for the research was submitted to the Bioethics Committee of the Faculty of Medicine of the Autonomous University of Querétaro, which approved it without any observation.

### Blood sampling

A sample of 5 ml of blood was extracted from the ulnar vein of each infant, using test tubes (Vacuttainer, Beckton-Dickinson). The serum samples were kept at 4 °C until the diagnostic tests were applied. We worked with a negative control serum, obtained from 20 healthy humans, who had not received blood transfusion, nor organ transplantation, nor had lived in a country endemic to Chagas disease. These samples were obtained in the CETS (State Blood Transfusion Center) in the State of Querétaro.

### Extraction and purification of excreted SOD (SODe)

**Cell culture:** As a biological material for the study and characterization of SOD, epimastigote forms of *Trypanosoma cruzi*, of the Cali strain, of Colombian origin, were used and perpetuated by consecutive reseeding in the laboratory. This strain has remained infective,

inoculating it every six months to a golden hamster from Syria (*Mesocricetus auratus*), recovering after a period of twenty days highly infective trypomastigote forms, which were again adapted to culture.

The parasite was cultured *in vitro* at 27 °C in Cellstar® tissue culture flasks (Greiner Bio-One®), using Grace's Insect Medium (Gibco®) supplemented with 10% V/V of fetal bovine serum (SBF, PAA®), previously inactivated by heating (56 °C, 30 min) to a growth density (estimated by counting in a hemocytometric count chamber of Neubauer®) of approximately 10<sup>7</sup> cells/mL. The flagellates were collected by centrifugation (1500 g/10 min) in the exponential phase of growth, following the methodology described by [19]. The sediment or pellet thus obtained was resuspended in Grace's medium not supplemented with SBF and cultured in the same conditions for 24-36 h, time after which the trypanosomatids were again collected by centrifugation, decanting the supernatant and keeping separately both fractions (S<sub>1</sub> and P) for the purification stages.

**Preparation of cellular extract:** The fractions resulting from the centrifugation (S<sub>1</sub> and P) were processed independently in order to purify both the total SOD activity and the excreted SOD (SODe), respectively.

First, the flagellates pellet obtained (around 0.5-0.6 grams wet weight) was subjected to a lysis or cell breakage process. For this, once the remains of the culture medium were eliminated by means of two washes with phosphate buffer (phosphate buffer saline, pH 7), said sediment was resuspended in 3 mL of buffer buffer STE Buffer 1 (250 mM sucrose, 25 mM Tris-HCl pH 7.4 and 1 mM EDTA) and cold sonicated in three cycles of 60 V and 30 s (with intervals of 1 min between cycles).

**Purification of SOD:** The sonicate was centrifuged (2500 gravities/10 min/4 °C) to discard the remains of membranes and dead cells, obtaining a new supernatant that we call Homogenate (fraction H).

In parallel, the initial supernatant (S<sub>1</sub>), which contains the cell excretion products, was filtered in order to retain the residues that have escaped the centrifugation using a 0.2 µm pore size microfilter (Minisart®, Sartorius®) (Fraction F).

Subsequently, parasites in MTL medium without fetal serum were cultured for 24 hours; the supernatant was collected by centrifugation at 600 gravities for 10 minutes and then passed through a 0.45 µm pore filter and solid ammonium sulfate was added. The protein fraction, which precipitated between 35% and 85% salt concentration, was centrifuged (9000 gravities for 20 min at 4 °C), dissolved in 2.5 ml of 20 mM potassium phosphate buffer (pH 7.8) containing 1 mM EDTA and desalted on a Sephadex G -25 column (Pharmacia, PD10), previously balanced with Buffer 2, bringing it to a final volume of 2.5 mL (SODe fraction) [20].

The fraction obtained from SODe was used as an antigen in the ELISA and Western Blot assays. The protein content was determined using a Bio-Rad test, based on the Bradford method (Sigma-Aldrich, St. Louis, M.O.), with bovine serum albumin as a standard.

### Serological test (ELISA)

The purified protein fraction (SODe) at a concentration of 1.5 µg was coated onto polystyrene microtiter plates (Nunc, Denmark) in carbonate buffer (pH 8.2) for 2 hours at 37 °C. The antigen stuck on the plate was removed by washing three times with PBS-Tween 20, 0.05% (washing buffer). The free adsorption sites were taken by incubation (2 hours at 37 °C) with blocking buffer (PBS-Tween 20, 0.2%, BSA 1%). Once washed as described above, the plates were incubated (45 minutes at 37 °C) with a serum dilution of 1/80 in wash buffer. After a second wash, the plates were incubated in the dark for 20 minutes with 100 µL of an antibody conjugated with enzyme (anti-IgG human peroxidase, Sigma) at a dilution of 1:1000. The reaction of the enzyme was catalyzed by the application of the chromogenic substrate OPD (o-phenylenediamine dihydrochloride, Sigma) and 10 µL of 30% H<sub>2</sub>O<sub>2</sub> in 25 mL for 20 minutes in the dark. The reaction was then stopped by the addition of 50 µL of 3N HCl.

Absorbance was read at 492 nm in microplate reader (Sunrise, TECAN). All samples were analyzed in triplicate in polystyrene microtiter plates. The mean and standard deviations (SD) of the optical densities of the negative control sera (20 healthy humans) were applied to calculate the cut-off value (mean + 3 × SD) [15].

### Analysis of western blot

The SODe antigen fraction (at a concentration of 1.5 µg protein) was in IEF 3-9 gels and then transferred to a nitrocellulose membrane (Hybond C Extra, Amersham Pharmacia Biotech) using the Phast-Transfer kit, as described by the manufacturer (Phast-System Manual). The membrane was blocked for 2 hours at room temperature using 0.4% gelatin and 0.2% Tween 20 in PBS, followed by three washes, using 0.1% Tween 20 in PBS solution (PBS-T) and incubated for 2 hours at room temperature, with donor serum at a 1/100 dilution. Before washing, the membrane was further incubated for 2 hours at room temperature with the second antibody, anti-human immunoglobulin G (Fc-specific) peroxidase conjugate (Sigma Immunochemical, 1/1000 dilution). After washing as indicated above, diaminobenzidine (0.5 mg/ml in 0.1 M Tris/HCl buffer, pH 7.4, containing 1/5000 H<sub>2</sub>O<sub>2</sub> [10 v/v]) was added and the reaction stopped with several washes in distilled water [15].

### Statistic analysis

The results obtained in the serological study for each chosen variable were processed in the Excel Program

for the elaboration of the Tables. For the statistical correlation between the ELISA-SODe and WB-SODe tests, the SPSS version 11.5 program for Windows was used, calculating the parameters of Sensitivity, Specificity, Negative Predictive Value (NPV), Positive Predictive Value (PPV), Index of Kappa and Concordance Percentage.

### Results

Of the total of 540 children samples analyzed with the ELISA-SODe and WB-SODe techniques, 106 were reactive to one or another test, 99 for ELISA-SODe for *T. cruzi* (18.3%) and 100 reactive sera for WB-SODe (18.52%) (Table 2).

The SPSS program, version 11.5, showed a sensitivity of 94%, specificity of 98%, PPV was 98% and NPV was 98%. The Kappa index presented a value of 0.95, which confirmed the agreement, between both tests, ELISA-SODe and WB-SODe (Table 3).

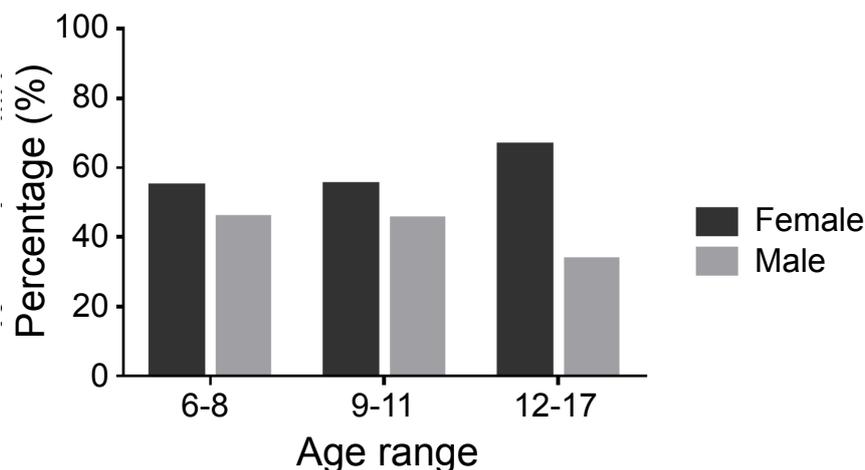
**Table 1:** List of sera studied in rural and urban pediatric population of the state of Querétaro (Mexico), by location, gender and age.

	N° Sera	Total
<b>Male population</b>		
Rural Male Population	148	
Urban Male Population	117	
		<b>265</b>
<b>Female population</b>		
Rural Female Population	148	
Urban Female Population	127	
		<b>275</b>
<b>Total</b>		
		<b>540</b>
<b>Population of 6 to 8-years-old</b>		
Rural Male Population	60	
Urban Male Population	52	
Rural Female Population	67	
Urban Female Population	60	
		<b>239</b>
<b>Population of 9 to 11-years-old</b>		
Rural Male Population	64	
Urban Male Population	54	
Rural Female Population	58	
Urban Female Population	56	
		<b>232</b>
<b>Population of 12 to 17-years-old</b>		
Rural Male Population	24	
Urban Male Population	11	
Rural Female Population	23	
Urban Female Population	11	
		<b>69</b>
<b>Total</b>		
		<b>540</b>

**Table 2:** Relationship of reactive sera from pediatrics (rural and urban population) of Queretaro (Mexico) by ELISA and Western Blot using SODe antigen fractions of *T. cruzi* associated with age and gender.

Sera	Age	Sex	ELISA-SODe <sup>a</sup>	WB-SODe <sup>b</sup>	Sera	Age	Sex	ELISA-SODe <sup>a</sup>	WB-SODe <sup>b</sup>	Sera	Age	Sex	ELISA-SODe <sup>a</sup>	WB-SODe <sup>b</sup>	Sera	Age	Sex	ELISA-SODe <sup>a</sup>	WB-SODe <sup>b</sup>
5	7	M	+	+	169	6	F	+	+	321	10	M	+	-	435	10	F	+	+
29	6	F	+	+	172	7	M	+	+	335	6	M	+	+	447	7	F	+	+
37	9	M	+	+	176	11	F	+	+	336	6	M	-	+	449	7	M	+	+
38	11	M	+	+	178	13	M	+	+	338	6	F	-	+	450	6	F	+	+
48	8	F	+	+	179	11	M	+	+	341	8	M	+	+	453	8	F	+	+
59	14	F	+	+	186	11	F	-	+	344	6	F	+	+	458	11	F	+	+
61	15	M	+	+	193	6	F	-	+	345	7	M	+	+	459	9	F	+	+
67	14	F	+	+	214	8	M	+	+	351	6	M	+	+	464	6	F	+	+
68	15	F	+	+	216	10	F	+	+	353	6	M	+	+	475	6	F	+	+
85	8	M	+	+	242	11	F	+	+	355	8	M	+	+	477	7	F	+	+
90	6	F	+	-	244	11	F	+	+	357	7	M	+	+	488	9	F	+	+
92	7	F	+	+	245	10	M	+	+	360	7	M	+	+	491	8	F	+	+
96	6	F	+	+	248	7	M	+	-	370	8	F	-	+	493	8	F	+	+
97	6	F	+	+	249	7	F	+	+	380	9	F	+	+	502	10	M	+	+
98	6	F	+	+	250	6	M	+	-	387	11	M	+	+	504	10	F	+	+
100	8	M	+	+	267	8	M	+	+	388	9	M	+	+	510	10	F	+	+
102	10	F	+	+	272	9	F	+	+	392	9	M	-	+	513	10	M	+	+
108	9	F	+	+	275	8	F	+	+	403	10	M	+	+	518	12	M	+	+
120	6	F	+	+	277	8	F	+	+	404	11	F	+	+	519	12	M	+	-
128	6	M	+	+	278	9	F	+	+	409	10	F	+	+	526	11	M	+	+
132	7	M	+	+	287	8	M	+	+	415	12	F	+	+	531	12	F	+	+
135	11	M	+	+	288	10	M	+	-	416	8	M	+	+	532	12	F	+	+
136	11	M	+	+	290	11	M	+	+	418	9	F	+	+	537	10	M	+	+
142	6	F	+	+	301	8	M	+	+	421	10	M	+	+	539	9	M	+	+
151	6	F	+	+	304	8	F	+	+	427	10	F	+	+	540	9	M	+	+
156	11	M	+	+	305	8	F	+	+	431	10	F	+	+					
167	9	F	+	+	311	11	M	+	+	433	10	F	+	+					

<sup>a</sup>: ELISA-SODe: Enzyme-Linked Immunosorbent Assay (ELISA) using excreted superoxide dismutase (SODe) by epimastigotes of *T. cruzi* as antigen fraction; <sup>b</sup>: WB-SODe: Western Blot (WB) using excreted superoxide dismutase (SODe) by epimastigotes of *T. cruzi* as antigen fraction.



**Figure 1:** Percentages of positive sera in rural and urban pediatric population by gender in Queretaro, Mexico. The data were analyzed by Chi-square test.

Odds ratio = 0.9213. 95% Confidence interval 0.6563 to 1.293.

**Table 3:** Evaluation of the reliability to detect *T. cruzi* antibodies, using ELISA technique with SODe CRU antigen, in 540 sera of children from the state of Queretaro (Mexico). The values given are estimated with Western Blot as the comparator.

	ELISA/SODe CRU
Sensitivity	94%
Specificity	98%
Positive predictive value	98%
Negative predictive value	98%
Kappa index	0.95

The concordance between the two tests (ELISA-SODe and WB-SODe) established the positivity of 93 children, a prevalence of 17.22% of the total population. In the analysis of the results on the prevalence of the samples coming from rural and urban areas, it only showed a slight difference; Infection rates were higher (18.58%) in rural areas than in urban areas (15.98%) (Figure 1).

By age groups, in the total population studied, the prevalence of age range of 6 to 8 years was 20.08% and 9 to 11 years of 17.6%, and 13.04% of children aged 12 to 17 years. In rural areas, groups of children aged 6 to 8 years presented 21.2% seroprevalence, the group of 12 to 17 years, 13.04%. The results of the urban areas were reversed in comparison with the rural areas, where the groups with the highest values were those with ages of 12 to 17 years (18.2%), (Table 1) while children aged 6 to 8 years showed a lower seroprevalence of 14.3%.

The prevalence was slightly higher in women, 18.0% versus 16.6% in men in the total population. Therefore, no significant differences were observed by gender in the rural and urban areas that have the highest seroprevalence in women of 19% and men of 12% (Figure 2).

## Discussion

Due to the high incidence of Chagas disease in a wide

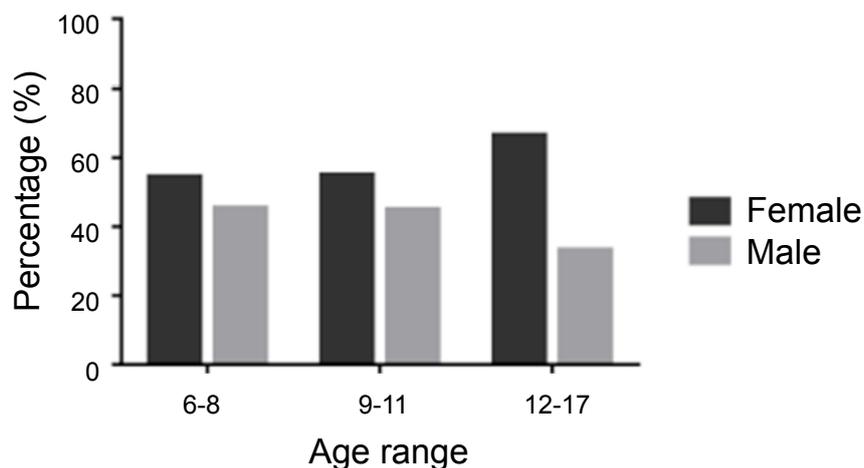
variety of nearby endemic areas surrounding Querétaro, such as Hidalgo, Michoacán, Guanajuato, San Luis Potosí and even the State of Mexico, (Figure 3), and the internal risk posed by humans and other mammalian species to act as natural reservoirs of *T. cruzi*, it is of the utmost importance that appropriate prevention measures are provided in the populations most at risk. It is crucial to keep updated the techniques of diagnosis and it is vital to develop highly sensitive and specific diagnostic methods to provide the appropriate treatment and in time, to reduce transmission of any kind.

In this sense, the present study proposes a confirmatory approach, since the use of the enzyme SODe used as antigen in ELISA and Western Blot, is highly effective for diagnose *T. cruzi* infections in human populations living in endemic areas of the disease.

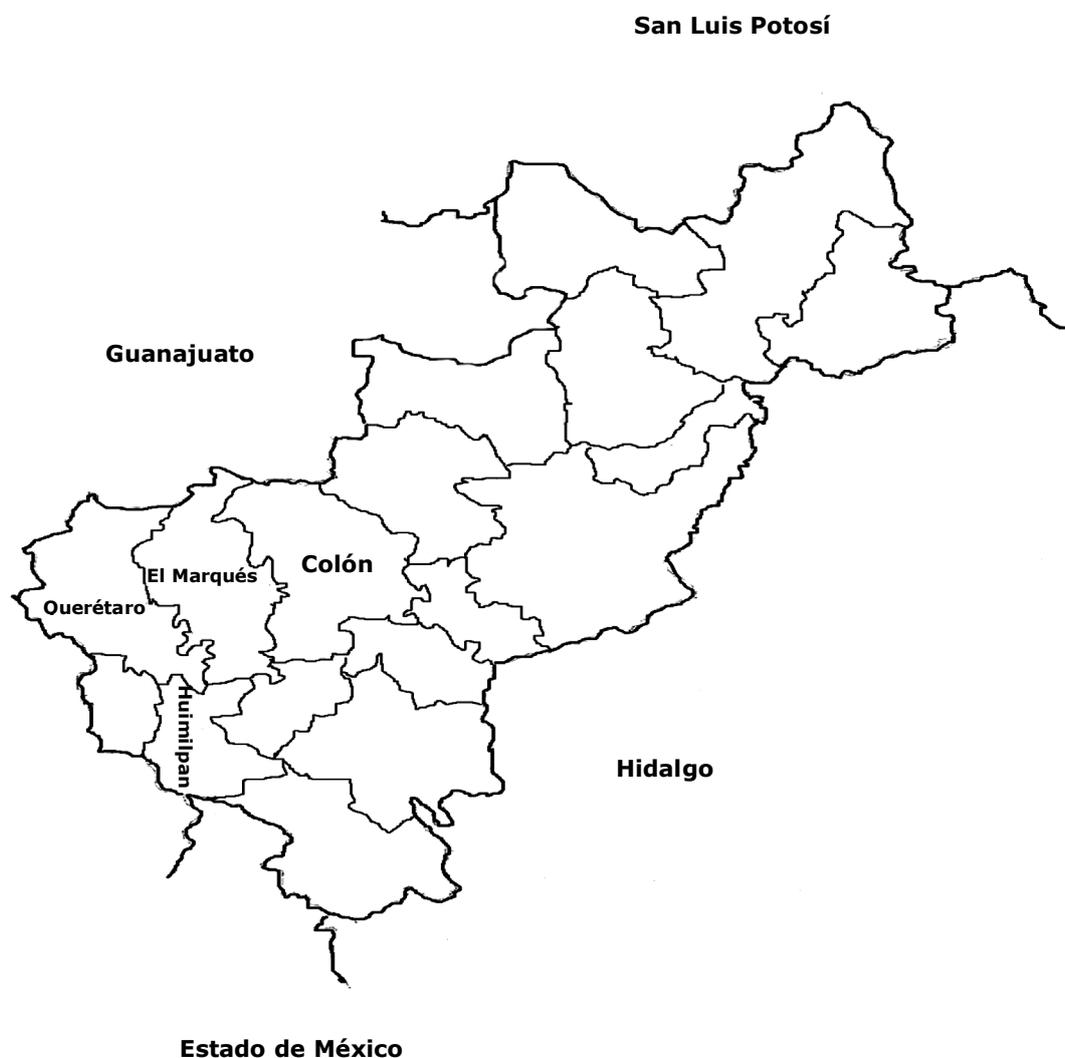
In addition, the presence of false positives with the ELISA technique may be due to cross reactions with other protozoa, mainly with *T. rangeli* and with species belonging to the genus *Leishmania spp*, among others. This is a serious inconvenience for patients from geographical areas that harbor other infectious agents.

The FeSOD enzyme has been identified in several protozoa as: *Plasmodium spp* [21] *Acanthamoeba castellanii* [22], *Entamoeba histolytica* [23], *Toxoplasma gondii* [24], *Trichomonas vaginalis* [25].

Mateo and collaborators in 2008, purified and characterized four SOD isoforms present in *T. cruzi*, one of them, with Isoelectric Point (IP) around 3.8 and Molecular Weight around 25 kDa [26]. Subsequently, the same authors showed that this isoform excreted by the parasite had immunogenic characteristics, so when used as an antigen in ELISA and/or Western Blot tests, it showed that it does not cross react and that it is specific to genus and species or to other trypanosomatids [11,15,17].



**Figure 2:** Percentages of positive sera of pediatric population by age range in males and females in Querétaro, Mexico. The data were analyzed by Chi-square test. Odds ratio = 0.9818. 95% Confidence interval 0.4152 to 2.321 in age groups 6-8 and 9-11. Odds ratio = 0.6000. % Confidence interval 0.1328 to 2.711 in age groups 6-8 and 12-17. Odds ratio = 0.6111. % Confidence interval 0.1337 to 2.794 in age groups 9-11 and 12-17. Software GraphPad Prism (version 6, GraphPad Software, Inc.).



**Figure 3:** Map of the state of Querétaro. The municipalities studied and neighboring states are indicated, whose seroprevalences estimated by various studies were shown as endemic to Chagas disease.

The evaluation of the results obtained with the total sera analyzed with the ELISA-SODE technique and the

WB-SODE technique was corroborated with previous studies, whose values of specificity and sensitivity

were very good. López-Céspedes and collaborators in 2012, conducted a cross-sectional study in adults of the suburban area of the municipality of Querétaro, obtaining a seroprevalence of 11%. Villagrán, in 2005, initiated the study with the SOD, using it to detect anti-*Trypanosoma cruzi* antibodies in serum samples, from the general population of endemic communities up to that time, from the same state, with 8.16% seroprevalence, later, in 2009, a serological study was carried out with known conventional tests and a statistical evaluation was carried out among the same tests applied. And finally in 2014, a comparison was made between conventional tests already known and SOD, in sera from rural communities that had not been studied and only in the pediatric population.

The high rate of infection detected in children in rural areas may be the result of congenital transmission and/or high vector exposure. The results of urban areas compared to rural areas, the age group of 12 to 17 years, resulted in higher values. These percentages can be attributed to the cumulative and prolonged exposure to infected vectors, which increases with age. There is no significant difference in the results by age and gender in rural and urban areas.

The prevalence in urban areas was significantly higher in women according to what was reported by other authors in their research in Puebla, Puebla [27], and in Tamazunchale, San Luis Potosí [28]. These preliminary data revealed the degree of infection by *Trypanosoma cruzi* in urban areas of our country.

## Acknowledgment

The authors thank the parents and principals of the elementary schools, who kindly allowed us to enter their classrooms to take blood samples from their students to conduct the research. To Dr. Manuel Sánchez Moreno, head of the Molecular Biology of Parasites, of the Faculty of Sciences of the University of Granada. Spain, that we opened the doors of your laboratory to perform tests with Superoxide dismutase, and we would like to express our gratitude to E. Guerrero López for the technical assistance in the preparation of the culture media.

## Financial Support

Cuerpo Académico UAQ-CA-97 Investigación Clínica Molecular. PRODEP.

## References

- Gutiérrez-Sotelo O, Molina-Solís S, Calvo-Fonseca N, Romero Triana L, Esquivel Alfaro L, et al. (2007) Enfermedad de Chagas en pacientes con miocardiopatía dilatada idiopática en Costa Rica. *Acta Médica Costarricense* 49: 97.
- Rassi A Jr, Rassi A, Marín-Neto JA (2010) Chagas disease. *Lancet* 375: 1388-1402.
- Pan American Health Organization (2006) Quantitative estimation of Chagas disease in the Americas, OP5 /HDM/CD/425-0G.
- Salazar-Schettino PM, Bucio-Torres MI, Cabrera-Bravo M, de Alba-Alvarado MC, Castillo-Saldaña DR, et al. (2016) Enfermedad de Chagas en México de. *Revista de la Facultad de Medicina UNAM* 59: 6-16.
- Hotez PJ, Dumonteil E, Heffernan MJ, Bottazzi ME (2013) Innovation for bottom 100 million: Eliminating neglected tropical diseases in the Americas. *Adv Exp Med Biol* 764: 1-12.
- Bern C, Montgomery SP (2009) An estimate of the burden of Chagas disease in the United States. *Clin Infect Dis* 49: e52-e54.
- Tarleton RL, Gürtler RE, Urbina JA, Ramsey J, Viotti R (2014) Chagas disease and the London Declaration on Neglected Tropical Diseases. *PLoS Negl Trop Dis* 8: e3219.
- Guzmán-Bracho C (2001) Epidemiology of chagas disease in Mexico: An update. *Trends Parasitol* 17: 372-376.
- Jiménez-Cardoso E, Campos Valdéz G, Cortes-Campos A, de la Luz Sanchez R, Mendoza CR, et al. (2012) Maternal fetal transmission of *Trypanosoma cruzi*: A problem of public health little studied in México. *Exp Parasitol* 131: 425-432.
- Luquetti AO (2007) Diagnóstico de la enfermedad de Chagas. In: Rosas F, Vanegas D, Cabrales M, Enfermedad de Chagas. Sociedad Colombiana de Cardiología y Cirugía CardioVascular, Bogotá, Colombia, 25.
- Portela-Lindoso AA, Shikanai-Yasuda MA (2003) Chronic Chagas' disease: From xenodiagnosis and hemoculture to polymerase chain reaction. *Rev Saúde Pública* 37: 107-115.
- Muro A, López-Abán J, Ternavasio-de la Vega HG, Pérez-Arellano JL (2010) Infecciones por protozoos flagelados hemotisulares II. Enfermedad de Chagas. *Tripanosomosis africana*. *Medicine* 10: 3609-3705.
- World Health Organization (2010) Grupo de trabajo científico. Reporte sobre la Enfermedad de Chagas. Programa especial de Investigaciones y enseñanzas sobre enfermedades tropicales (TDR), patrocinado por UNICEF, PNUD/BANCO MUNDIAL/OMS.
- Villagrán ME, Marín C, Rodríguez-González I, De Diego JA, Sánchez-Moreno M (2005) Use of an iron superoxide dismutase excreted by *Trypanosoma cruzi* in the diagnosis of Chagas disease: Seroprevalence in rural zones of the state of Queretaro, Mexico. *Am J Trop Med Hyg* 73: 510-516.
- Mateo H, Sánchez-Moreno M, Marín C (2010) Enzyme-linked immunosorbent assay with purified *Trypanosoma cruzi* excreted superoxide dismutase. *Clin Biochem* 43: 1257-1264.
- Marín C, Sánchez-Moreno M (2010) Excreted/secreted antigens in the diagnosis of Chagas' disease. In: Jirillo E, Brandonisio O, Immune Response to Parasitic Infections. Volume 1, Bentham Science Publishers E-book, 10.
- López-Céspedes A, Villagrán E, Briceño Álvarez K, de Diego JA, Hernández-Montiel HL, et al. (2012) *Trypanosoma cruzi*: Seroprevalence detection in suburban population of Santiago de Queretaro (Mexico). *Scientific World Journal* 2012: 914129.
- Longoni SS, Marín C, Sauri-Arceo CH, López-Céspedes A, Rodríguez-Vivas RI, et al. (2011) An iron superoxide dismutase antigen-based serological screening of dogs indicates their potential role in the transmission of cutaneous Leishmaniasis and Trypanosomiasis in Yucatán, México. *Vector Borne Zoonotic Dis* 11: 815-821.

19. Marín C, Hitos AB, Rodríguez-González I, Dollet M, Sánchez-Moreno M (2004) *Phytomonas* iron superoxide dismutase: A possible molecular marker. *FEMS Microbiol Lett* 234: 69-74.
20. Villagrán ME, Sánchez-Moreno M, Marín C, Uribe M, De la Cruz JJ, et al. (2009) Seroprevalence to *Trypanosoma cruzi* in rural communities of the state of Queretaro (México): Statistical evaluation of tests. *Clin Biochem* 42: 12-16.
21. Dive Daniel, Sylvie Gratepanche, Helene Yera, Philippe Bécuwe, Wassim Daher, et al. (2003) Superoxide dismutase in *Plasmodium*: A current survey. *Redox Report* 8: 265-267.
22. Jung-Yeon Kim, Byoung-Kuk Na, Kyoung-Ju Song, Mi-Hyun Park, Yun-Kyu Park, et al. (2012) Functional expression and characterization of an iron-containing superoxide dismutase of *Acanthamoeba castellanii*. *Parasitol Res* 111: 1673-1682.
23. Tannich E, Bruchhaus I, Walter RD, Horstmann RD (1991) Pathogenic and nonpathogenic *Entamoeba histolytica*: Identification and molecular cloning of an iron-containing superoxide dismutase. *Mol Biochem Parasitol* 49: 61-71.
24. Brydges SD, Carruthers VB (2003) Mutation of an unusual mitochondrial targeting sequence of SODB2 produces multiple targeting fates in *Toxoplasma gondii*. *J Cell Sci* 116: 4675-4685.
25. Viscogliosi E, Delgado-Viscogliosi P, Gerbod D, Dauchez M, Gratepanche S, et al. (1998) Cloning and expression of an iron-containing superoxide dismutase in the parasitic protist, *Trichomonas vaginalis*. *FEMS Microbiol Lett* 161: 115-123.
26. Mateo H, Marín C, Pérez-Córdon G, Sánchez-Moreno M (2008) Purification and biochemical characterization of four iron superoxide dismutases in *Trypanosoma cruzi*. *Mem Inst Oswaldo Cruz* 103: 271-276.
27. Sosa-Jurado F, Mazariego-Aranda M, Hernández-Becerril, Garza-Murillo V, Cárdenas M, et al. (2003) Electrocardiographic findings in Mexican chagasic subjects living in high and low endemic regions of *Trypanosoma cruzi* infection. *Mem Inst Oswaldo Cruz* 98: 605-610.
28. Juárez-Tobías S, Vaughan G, Torres-Montoya A, Escobar-Gutiérrez A (2009) Seroprevalence of *Trypanosoma cruzi* among Teenek Amerindian residents of the huasteca region in San Luis Potosí, México. *Am J Trop Med Hyg* 81: 219-222.