



# Mycophenolate Mofetil-Induced Colitis in a Pediatric Heart Transplant Recipient

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## Abstract

Mycophenolate Mofetil (MMF) is a frequently used immunosuppressive medication in pediatric heart transplant (HT) patients. Although therapeutic monitoring of MMF has been an integral part of routine practice for many years, there is growing evidence that genetic variation in host can lead to adverse outcomes such as drug toxicities. Maintaining a reasonable balance between efficacy and toxicity in an individual patient remains one of the major challenges in pediatric HT recipients. Here we describe a pediatric HT recipient who has suffered MMF induced colitis despite acceptable therapeutic levels.

## Introduction

Heart transplantation (HT) is a well-established therapy for end-stage otherwise untreatable heart disease. In recent years, outcome of pediatric HT have steadily improved with current 5-year overall survival rates estimated at 83% [1]. Despite these advances, children who receive HT experience a significant morbidity and mortality. There are many unknown clinical factors for considerable uncertainty in their outcome. One of the factors could be genetic variation in the host. There is growing evidence that genetic variation leads to differences in immune response, response to therapies and susceptibility to adverse outcomes such as drug toxicities [2].

Mycophenolate mofetil (MMF) is the most commonly prescribed adjunctive maintenance immunosuppressive medication in pediatric HT at the present time, with the use of azathioprine being on the decline. MMF is a prodrug that is rapidly hydrolyzed to the active form mycophenolic acid (MPA). MPA is a reversible inhibitor of inosine monophosphate dehydrogenase (IMPDH), which catalyzes purine synthesis. Because activated lymphocytes are dependent on the de novo synthesis of purine nucleotides, IMPDH inhibition causes decreased B- and T-cell proliferation and decreased antibody production [3]. MPA preferentially binds to IMPDH isoform type II, expressed in active lymphocytes.

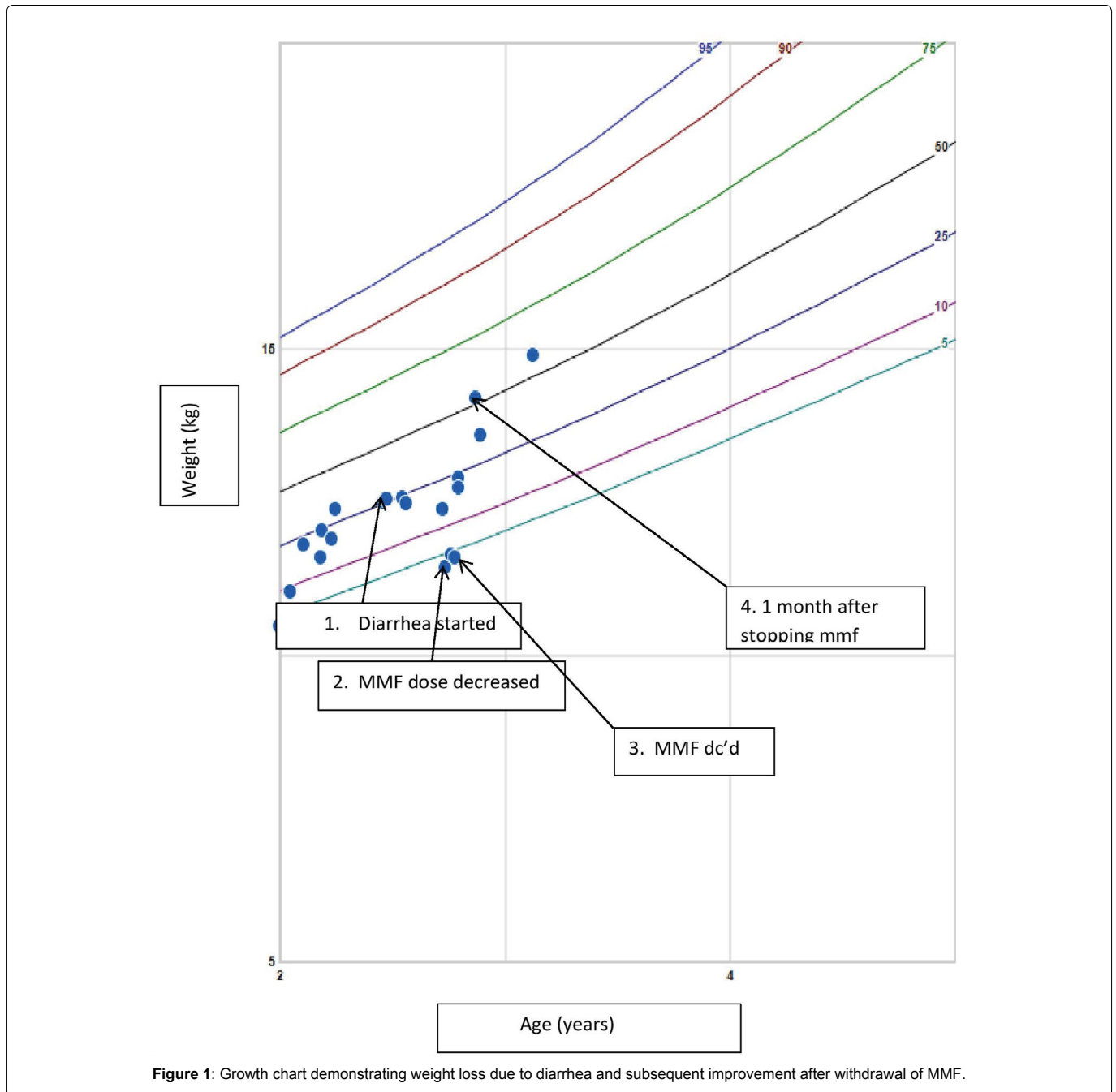
The most common adverse effects of MMF in pediatric heart transplant recipients include watery diarrhea, nausea, abdominal

cramping, and bone marrow suppression. MMF can lead to malignancy and infections (specifically CMV and herpes zoster infections), ulcerative esophagitis, reactive gastropathy, and graft-versus-host disease (GVHD)-like features in intestinal biopsies [4]. The MMF-related colitis is described in 2.7% adult renal transplant patients from a large series of 580 patients [5]. However, reports on the clinical course of colitis in pediatric heart transplant recipients are scarce. We describe a case of severe MMF induced colitis in a child 15 months after heart transplant.

## Case

Our patient is a 3 year old Caucasian boy who underwent orthotopic HT at 14 months of age for hypoplastic left heart syndrome and Glenn failure. His induction immunosuppression consisted of basiliximab and methylprednisone, and for maintenance tacrolimus, MMF, and prednisone. Prednisone was discontinued at 12 months after HT. His tacrolimus dose was adjusted per level to maintain trough between 5-10 ng/mL. MMF dose was adjusted for MPA level with a therapeutic range between 1 and 3.5 mcg/ml. He was negative for cytomegalovirus (CMV) and Epstein-Barr virus (EBV) before HT, and received a CMV negative and EBV positive donor heart. He remained on antimicrobial prophylaxis with trimethoprim-sulfamethoxazole as per our institutional protocol. His serial polymerase chain reaction (PCR) for EBV and CMV were negative after HT.

His first 15 months post-HT was uneventful. In the 16<sup>th</sup> month he entered a 3 month period of intermittent non-bloody diarrhea alternating with constipation and severe weight loss (Figure 1). There was no fever, and his physical examination was only significant for nonspecific abdominal tenderness. The gastrointestinal panel was negative for aeromonas, campylobacter, clostridium difficile toxin A/B, plesiomonas, shigelloides, salmonella, yersinia enterocolitica, *E. coli*, cryptosporidium, cyclospora cayetanensis, entamoeba histolytica, giardia lamblia, norovirus and rotavirus. The only positive tests were for adenovirus from his stool, and transient disaccharides deficiency. Subsequent stool and blood PCR for adenovirus were negative. Elimination of dairy products reduced frequency of diarrhea from



**Figure 1:** Growth chart demonstrating weight loss due to diarrhea and subsequent improvement after withdrawal of MMF.

12-15 times per day to 6-8 times. During this 3 month period, he was admitted to the hospital multiple times with recurrent vomiting, diarrhea, dehydration, and one episode of acute renal insufficiency. He underwent an esophagogastroduodenoscopy and a colonoscopy. His colon biopsy showed reactive changes with crypt dropout, regeneration, and focal active colitis extending from cecum to rectum and was attributed to MMF colitis (Figure 2A and Figure 2B). The colonic tissue PCR for CMV, EBV and adenovirus were negative. There was no evidence of post-transplant lymphoproliferative disease. His MMF was stopped and azathioprine was started as adjunctive therapy for maintenance immunosuppression. Diarrhea improved within a week after MMF withdrawal and patient started gaining weight (Figure 1). He remained asymptomatic for the last 12 months as of this report.

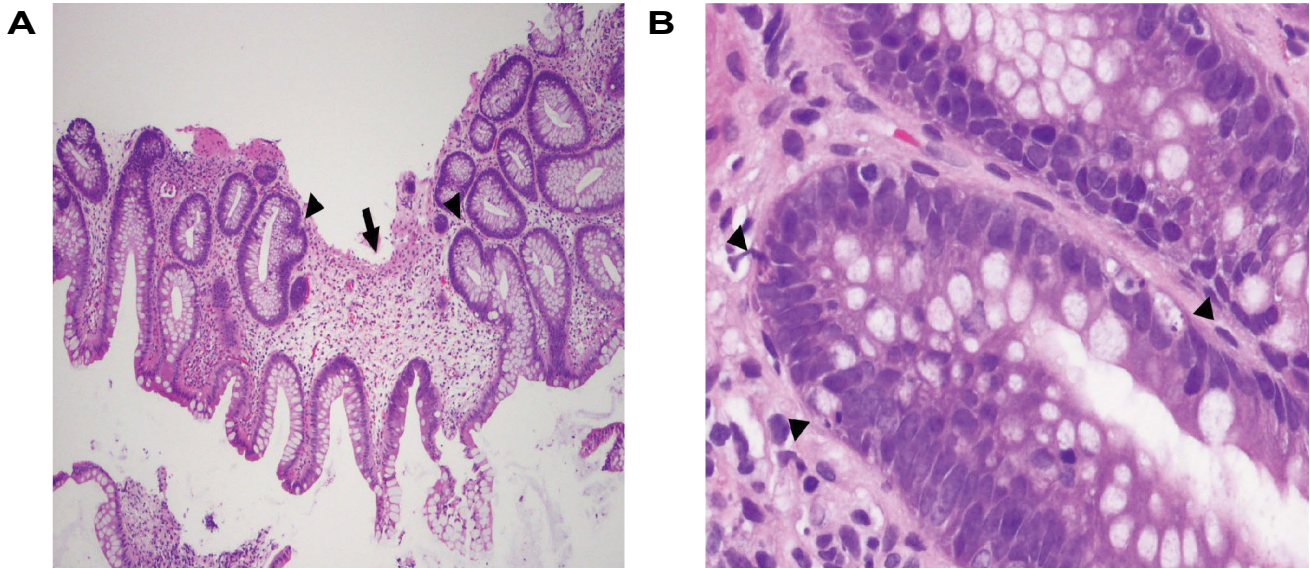
We monitored our patient every month with laboratory tests including cardiac biomarkers, end-organ functions and echocardiogram for allograft dysfunction. As our patient is clinically doing well, we did not repeat colonic biopsy nor re-challenged him with MMF. Our plan in future is if we see any evidence of either rejection or development of de novo donor specific antibodies, we will optimize his immunosuppression by reintroducing MMF

instead of Azathioprine. We also plan to evaluate this patient for single nucleotide polymorphism (SNP) associated with genetic polymorphism and drug toxicities if we need to resume MMF in future.

### Discussion

Recently, a case of late onset severe colitis attributed to MMF was described in an adult HT patient [6]. Pathak et al. [7] described a case of MMF induced colitis in a child after renal transplant which was confused with inflammatory bowel disease (IBD) [7]. Papadimitriou et al. [8] have described marked similarities in the histologic features of the MMF colitis and GVHD in children [8]. The common histopathological findings of MMF colitis and GVHD are prominent crypt enterocyte apoptosis accompanied by degenerative and regenerative changes (Figure 2A and Figure 2B).

The pathogenetic mechanism through which MMF exerts its toxicity in the gastrointestinal tract is unknown. One of the proposed mechanisms by which MPA could lead to diarrhea is inhibition of the de novo purine synthesis pathway, thereby preventing gastrointestinal epithelial cell growth and replication [9]. It has been suggested that an MMF dose of > 600 mg/m<sup>2</sup> every 12 hours is associated with



**Figure 2A & B:** 2A: Colonic mucosa showing crypt dropout (arrow) and regenerative branching crypts (arrowheads)  
2B: Colonic mucosa (high power) showing apoptosis of crypt cells (arrowheads)

higher plasma levels of MPA, and may be associated with a higher incidence of gastrointestinal side effects [10]. Our patient was on a dose of 740 mg/m<sup>2</sup> every 12 hours and his MPA level was between 1-2 mcg/mL without any evidence of bone marrow suppression such as leucopenia or anemia, suggesting that the mechanism of MMF induced gastrointestinal toxicities could be multifactorial.

The pharmacogenomics study of MMF in 59 pediatric HT patients has shown that ABCC2 rs717620 GG phenotype was protective against gastrointestinal side effects due to MMF. The protective mechanism of this phenotype is attributed to decreased enterohepatic recirculation and lower intestinal MPA concentrations [11]. MPA is metabolized through phase 2 glucuronidation by UDP glucuronosyltransferases (UGTs). In pediatric renal transplant patients and adult HT patients, polymorphisms in UGT2B7 and UGT1A8 have influenced the metabolism, clearance and side effect profile of MMF [12-14].

MMF-induced colitis should be considered in the differential diagnosis of patients taking this drug. The discontinuation of MMF resulted in good clinical outcome with resolution of diarrhea and no evidence of allograft rejection in our patient. There was no correlation of MMF-induced colitis with MPA level in our patient and might suggest individual pharmacogenetic variations that may influence drug toxicities. In future, pharmacogenomics study of MMF as a routine clinical practice may provide opportunity to minimize rejection events while avoiding serious toxicities in pediatric heart transplant recipients.

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