



ORIGINAL ARTICLE

Transforming Growth Factor-Beta (TGF- β) Regulates Nerve Growth Factor and Cyclooxygenase-2 Expression in Subsynovial Connective Tissue in Patients with Carpal Tunnel Syndrome

Mitsufumi Nakawaki¹, Kentaro Uchida^{1*}, Kenji Onuma¹, Koji Sukegawa¹, Toshihide Matsumoto², Gen Inoue¹, Hiroyuki Sekiguchi³ and Masashi Takaso¹

¹Department of Orthopedic Surgery, Kitasato University School of Medicine, Sagamihara City, Japan

²Department of Pathology, Kitasato University School of Medicine, Sagamihara City, Japan

³Shonan University of Medical Sciences Research Institute, Chigasaki City, Japan

*Corresponding author: Kentaro Uchida, Department of Orthopedic Surgery, Kitasato University School of Medicine, 1-15-1 Minami-ku Kitasato, Sagamihara City, Kanagawa 252-0374, Japan



Abstract

Background: It has been reported that transforming growth factor-beta (TGF- β) is elevated in the subsynovial connective tissues (SSCTs) of patients with idiopathic carpal tunnel syndrome (CTS) and TGF- β induced fibrosis of SSCTs. A TGF- β -mediated pathway may be a potential target for CTS therapy. Cyclooxygenase-2 (COX-2) and nerve growth factor (NGF) are regulated by TGF- β and are associated with fibrosis in several tissues; however, regulation in SSCTs has not been fully clarified.

Methods: Forty-three SSCTs were sampled from 35 patients undergoing carpal tunnel release (CTR) surgery. Correlation between TGF- β mRNA and protein expression and COX-2 and NGF were evaluated using quantitative PCR (q-PCR) analysis of total RNA sampled from the SSCTs. To investigate regulation of COX-2 and NGF by TGF- β , cultured SSCT cells were stimulated with TGF- β and COX-2 and NGF mRNA and protein expression were evaluated using q-PCR and western blotting.

Results: TGF- β -, NGF-, and COX-2-positive cells were observed in SSCTs. Expression of COX-2 and NGF positively correlated with TGF- β mRNA expression in the SSCTs of CTS patients (COX-2, $r = 0.629$, $p < 0.001$; NGF, $r = 0.521$, $p < 0.001$). The expression of COX-2 and NGF mRNA and protein increased significantly in SSCT cells following exogenous treatment with TGF- β compared to vehicle-treated cells ($p < 0.05$).

Conclusions: TGF- β may regulate COX-2 and NGF in the SSCTs of CTS patients.

Keywords

Subsynovial connective tissues, Transforming growth factor- β , Nerve growth factor, Cyclooxygenase-2

Introduction

Carpal tunnel syndrome (CTS) is a common neuropathy encountered in orthopedics. It results from compression of the distal median nerve (MN) within the carpal tunnel. Characteristic complaints of CTS are numbness, paresthesia, and pain in the hand and fingers, often exacerbated at night [1]. This characteristic MN compression results in various degrees of disability [2,3]. In recent years, elucidation of the pathological condition has progressed; however, the mechanism is still unknown.

Transforming growth factor- β (TGF- β) plays an important role in fibrotic reactions [4]. TGF- β levels are increased in injured tissues, and excessive TGF- β production has induced fibrosis in several tissues [4,5]. In the wrist, Subsynovial connective tissue (SSCT) envelops the nine flexor tendons and the MN in the carpal tunnel. Recent studies have shown that increased fibrosis of SSCT contributes to CTS development and is involved in the median nerve compression [6-9]. Increased TGF- β expression has been observed in the SSCT of patients with idiopathic CTS [10] and fibrosis is associated with



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Table 1: Primers sequences in this study.

Primer	Sequence (5'–3')	Product size (bp)
COX-2-F	TGGCTGAGGGAACACAACAG	74
COX-2-R	AACAAGTGCATCACCCCA	
NGF-F	CCCATCCCATCTTCCACAGG	74
NGF-R	GGTGGTCTTATCCCCAACCC	
GAPDH-F	TGTTGCCATCAATGACCCCTT	202
GAPDH-R	CTCCACGACGTACTIONCAGCG	

elevated activity of TGF- β [9]. Therefore, TGF- β -mediated pathway may be important in CTS pathology, albeit that its mechanism is not fully understood.

Recent studies have reported that TGF- β stimulates molecules involved in pain, including cyclooxygenase-2 (COX-2) and nerve growth factor (NGF) in smooth muscle cells, [11] epithelial cells, [12] and the articular cartilage of osteoarthritis patients [13]. Notably, a range of studies have reported that COX-2 and NGF may contribute to liver and cystic fibrosis [14]. TGF- β -mediation of these pathways may be a potential target for CTS therapy; however, regulation of COX-2 and NGF by TGF- β in SSCTs has not been examined.

Here, we investigated TGF- β regulation of COX-2 and NGF using SSCT harvested from CTS patients.

Materials and Methods

Human recombinant TGF- β was obtained commercially from R&D Systems, Inc. (Minneapolis MN, USA). Rabbit polyclonal primary antibodies against COX-2 (cat. no. ab52237) and rabbit monoclonal primary antibodies against NGF (cat. no. ab52918) were purchased from Abcam (Cambridge, UK).

We performed an unregistered retrospective study to investigate the correlation between the gene expressions of SSCTs. This study received approval from the Ethics Review Board of Kitasato University (reference number: KMEO B13-113). Consent to participation was provided by the patients for collection of SSCTs for examination. Inclusion criteria included a clear diagnosis of CTS, a neural electrophysiological examination conducted in the outpatient department of our hospital, and subsequent treatment by carpal tunnel release (CTR) surgery (CTRS). A total of 35 patients with CTS underwent carpal tunnel release CTRS at our center. The study included 12 men and 23 women aged 38–87 years (mean \pm SD, 68.3 \pm 10.9 years) with a mean \pm SD body mass index (BMI) of 24.0 \pm 5.8 kg/m² (range 13.6–44.0). A sample of 43 SSCT was harvested from each operated hand during the CTR. A portion of 35 SSCT tissue samples was kept at -80 °C until use in RNA extraction for real-time PCR analysis. SSCT from eight patients were additionally utilized for cell culture. The remaining tissue samples were used for immunohistochemistry. Each patient provided their informed consent to inclusion in the study the day preceding surgery.

Total RNA was obtained from the sampled SSCT by

use of TRIzol reagent (Invitrogen, Carlsbad CA, USA) as described previously [15,16]. cDNA was synthesized using SuperScript III RT (Invitrogen) in accordance with the manufacturer's protocol. The primers were developed by use of the Primer Blast software and synthesized at Hokkaido System Science Co., Ltd. (Sapporo, Japan). PCR primer pair sequences are provided in Table 1. Specificity of the amplified products was evaluated with the use of melt curve analysis, while quantitative PCR was conducted with the Real-Time PCR Detection System (CFX-96; Bio-Rad, Richmond CA, USA) to determine relative levels of mRNA expression. mRNA expression was normalized against GAPDH mRNA levels.

To investigate whether TGF- β regulates COX-2 and NGF expression in SSCT, cells from the SSCTs of six CTS patients were isolated from SSCT by digestion of the tissue with 5 mL of 1 mg/mL type I collagenase solution. The obtained cells were cultured in minimum essential medium alpha (α -MEM) supplemented with 10% fetal bovine serum in six-well plates. After 7 days, the cells were exposed to 1 and 10 ng/mL human recombinant TGF- β for 24 h. Controls were exposed to culture medium without TGF- β . Following treatment, the cells were collected for isolation RNA (see above), and COX-2 and NGF expression was evaluated using RT-PCR. Cells were also collected and subject to protein extraction (described below), and COX-2 and NGF protein expression was analyzed by western blotting.

To investigate COX-2 and NGF protein expression, SSCT cells stimulated with 1 or 10 ng/mL TGF- β were lysed in radioimmune precipitation buffer (RIPA) (Wako Pure Chemical, Tokyo, Japan) supplemented with a protease inhibitors (Sigma-Aldrich, MO, USA). Protein concentrations of the tissue extracts were then measured by a commercial bicinchoninic acid assay kit (Pierce, Rockford Illinois, USA). Protein extracts (10 μ g/lane) were then separated using sodium dodecyl sulfate-polyacrylamide gel electrophoresis. They were then electrophoretically transferred to a polyvinyl difluoride membrane and blocked for 1 h with a blocking reagent (Immunoblock, DS Pharma Biomedical Co., Ltd., Osaka, Japan). These blocked membranes were then incubated with rabbit primary antibodies against COX-2 and NGF at 4 °C overnight. The primary antibodies were diluted 1:1000 with blocking reagent. The membranes were then washed in tris-buffered saline (TBS-T) followed by incubation using horseradish peroxidase (HRP)-conjugated anti rabbit IgG antibodies (GE Healthcare, NJ, USA), following dilution at 1:1000 with blocking reagent. Immunoreactive proteins were visualized based on chemiluminescence using ECL substrate (Bio-Rad, Hercules, CA, USA), and images were visualized using X-ray film.

To reveal a normal distribution for this data, the Kolmogorov-Smirnov test were performed. Based on the results of this test, the relationship between TGF- β and COX-2 and NGF was evaluated using Pearson's

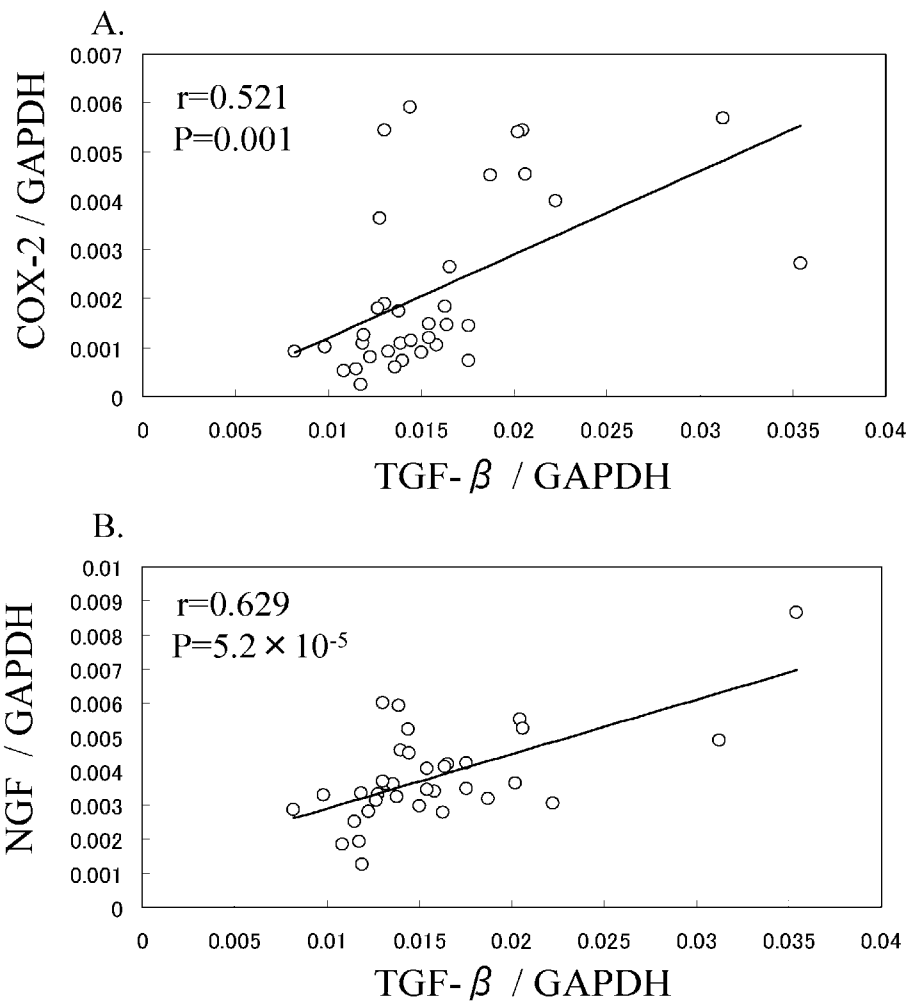


Figure 1: Correlation between mRNA expression levels of TGF-β and COX-2 and NGF in subsynovial connective tissue. Correlation between TGF-β and COX-2 (A) and NGF (B) mRNA expression levels in subsynovial connective tissue harvested from 35 CTS patients. *Pearson's coefficient $p < 0.05$.

correlation coefficient. A $p < 0.01$ was held to indicate statistical significance in correlation coefficient analysis. Potential statistical outliers that might have influenced linear regression coefficient analysis were identified with Cook's distance statistical test. Following the F-test, while differences between the vehicle control and TGF-β-treated SSCT cells were compared via one-way ANOVA using the Fisher least significant difference test. A $p < 0.05$ was considered statistically significant. All statistical analyses were conducted using SPSS (v. 19.0; SPSS, Chicago IL, USA).

Results

The expression levels of TGF-β were positively correlated with COX-2 (Figure 1A, $r = 0.521$, $p < 0.001$). Correlation between the TGF-β mRNA expression and NGF mRNA expression levels were also detected (Figure 1B, $r = 0.629$, $p < 0.001$).

The effects of TGF-β on the mRNA and protein expression of COX-2 and NGF were next examined. Real-time PCR analysis indicated that the gene expression of COX-2 and NGF was significantly raised in synovial cells in the presence of exogenous TGF-β in comparison

with vehicle control cells but was (Figure 2A and Figure 2B). COX-2 and NGF protein expression also increased with exposure to TGF-β (Figure 3).

Discussion

In this study of the mechanism of NGF and COX-2 regulation by TGF-β in the SSCT of CTS patients, we detected correlations between TGF-β concentration and NGF and COX-2 expression. Further, treatment cultured SSCT cells with TGF-β increased the expression of NGF and COX-2. This suggests that TGF-β regulates COX-2 and NGF in SSCT of CTS patients and may contribute to CTS pathology.

COX-2 upregulation was observed in CTS patients in the absence of inflammatory reactions and COX-2 mRNA expression in the synovium of CTS patients correlated with synovial hypertrophy [17]. Recent studies suggested that TGF-β expression may be a therapeutic target for the therapy of the non-inflammatory fibrosis developed in CTS since its expression is increased [10,18]. Previous studies also reported that COX-2 was regulated by TGF-β in granulosa cells, [19] fibroblasts, [20] and epithelial cells [12]. Hui, et al. reported that

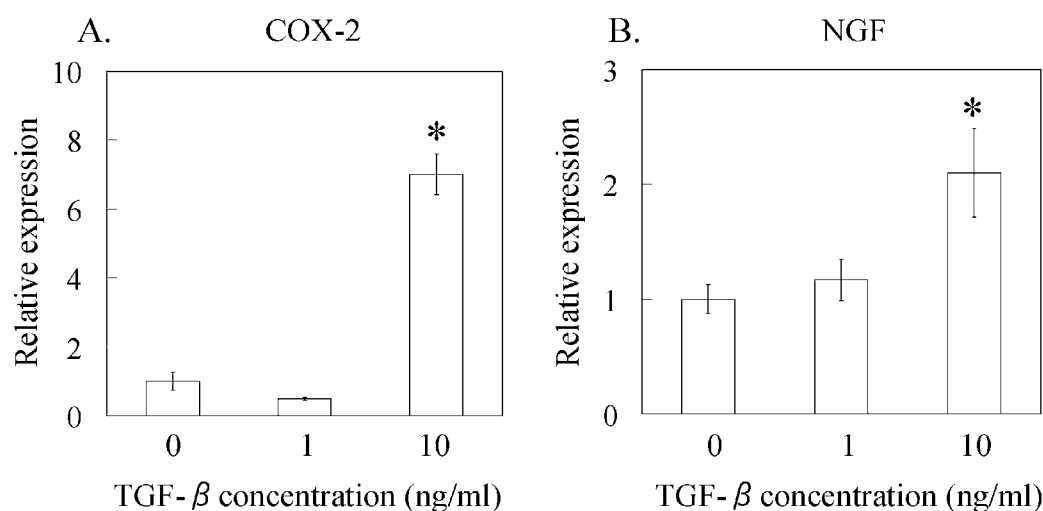


Figure 2: Effect of TGF- β on COX-2 and NGF mRNA expression in subsynovial connective tissue cell culture. Real-time polymerase chain reaction analysis for (A) cyclooxygenase-2 (COX-2) and (B) Nerve growth factor (NGF).

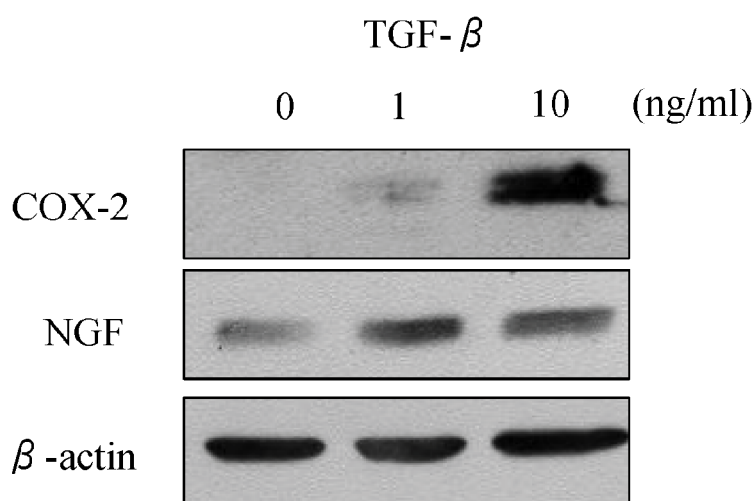


Figure 3: Effect of TGF- β on COX-2 and NGF protein expression in subsynovial connective tissue cell culture. Western blotting analysis for COX-2 and NGF. Subsynovial connective tissue cells were stimulated with 1 and 10 ng/ml TGF- β for 24 h prior to protein extraction and analysis of COX-2 and NGF protein.

TGF- β stimulates fibrosis via promotion of COX-2/PGE2 pathway in hepatic stellate cells [21] and that COX-2 inhibition reduced hepatic fibrosis. Here, there was a correlation between TGF- β and COX-2, and exogenously added TGF stimulated COX-2 mRNA and protein expression in SSCT cells. Based on previous studies, our results suggest that TGF- β -mediated COX-2 elevation may contribute to fibrosis in SSCTs of CTS patients.

Several studies reported that NGF production and release were promoted by inflammatory cytokines, which include tumor necrosis factor-alpha (TNF- α) and interleukin-1-beta (IL-1 β) under inflammatory or non-inflammatory conditions by TGF- β , and that elevation of NGF contributes to musculoskeletal pain [13,22-24]. For example, TNF- α and IL-1 β promoted NGF mRNA expression in synovial cell culture [25]. TGF- β stimulated NGF expression in osteoarthritic chondrocytes [13]. In contrast, NGF may exacerbate fibrosis in tissues such as

liver, lung, and skin [25-27]. Micera, et al. reported that NGF promotes migration of lung and skin fibroblast and could exert a pro-fibrogenic effect [26]. NGF expression in hepatocytes during liver injury acts as a paracrine mediator for hepatic satellite cell-induced fibrosis [27]. Here, there was a correlation between TGF- β and NGF, and exogenously added TGF stimulated NGF at both mRNA and protein level in SSCT cells. Our results suggest that NGF is also regulated by TGF- β and may contribute to fibrosis in SSCTs of CTS patients.

Various clinical and clinical trials have demonstrated that COX-2 inhibitor and anti-NGF antibody show therapeutic efficacy in the treatment of pain in musculoskeletal disorders [28-31]. The non-steroidal anti-inflammatory drugs (NSAIDs) containing COX-2 inhibitors are among the most widely used classes of drugs for the acute and chronic pain management for musculoskeletal disorders [28]. NGF neutralization antibody also

has also robust analgesic effects in osteoarthritis (OA) [30,31] and low back pain [29,31]. Clinical trials of tanezumab in human OA patients and patients with low back pain are underway. In the present study, although we did not determine the association of COX-2 and NGF expression levels and pathology in CTS patients, findings concerning TGF- β -mediated COX-2 and NGF regulation in SSCTS may be a target for future treatments of CTS.

Several limitations of our study deserve mention. First, the absence of a control, non-CTS patient population reduces the certainty of our results. The question of whether COX-2 and NGF levels are elevated in the SSCT of CTS patients as compared to non-CTS patients requires clarification. Second, histological examination of fibrosis and edema are needed to reveal the causative CTS pathology. Finally, we did not determine if a causative link exists between pathology and COX-2 and NGF in CTS patients.

Conclusions

TGF- β may regulate COX-2 and NGF in the SSCTs of CTS patients.

Conflicts of Interest

None to declare.

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